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Research article



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Productivity and immune reactivity of small cattle with different genotypes

ABSTRACT

Modern methods of breed formation in animal husbandry underscore the importance of studying the influence of genetic factors on the productive qualities of farm animals. This research analyzes the association between candidate gene polymorphisms and meat productivity indicators and immune status in sheep. Genotyping of the GH, GDF9, and CAST loci was performed using the PCR-RFLP method in a sample of Dagestan Mountain sheep. Statistically significant associations were established between the genetic variants and the traits under study. Animals with the GH/HaeIII AB and GDF9/AspEI AA genotypes were characterized by an increased level of CD4⁺ lymphocytes and a reduced content of CD8⁺ cells, resulting in higher values of the immunoregulatory index compared to carriers of the GH/HaeIII AA and GDF9/AspEI GG genotypes. Analysis of productive qualities revealed the advantage of the CAST MM genotype over CAST MN in terms of live weight. A similar pattern was established for the GH AA genotype compared to GH AB. The obtained data demonstrate the promise of using molecular genetic markers in breeding programs for the simultaneous improvement of both productive characteristics and the immune status of sheep.

Key words: marker selection, genotyping, small ruminants, immune reactivity, productivity

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Научная статья



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Продуктивность и иммунная реактивность мелкого рогатого скота разных генотипов

РЕЗЮМЕ

Современные методы породообразования в животноводстве актуализируют задачу изучения влияния генетических факторов на продуктивные качества сельскохозяйственных животных. В рамках настоящего исследования проведен анализ взаимосвязи полиморфизма генов-кандидатов с показателями мясной продуктивности и иммунного статуса у овец. Методом ПЦР-ПДРФ осуществлено генотипирование по локусам GH, GDF9 и CAST в выборке горных дагестанских овец. Установлены статистически значимые ассоциации между генетическими вариантами и изучаемыми признаками. Животные с генотипами GH/HaeIII AB и GDF9/AspEIAA характеризовались повышенным уровнем CD4⁺-лимфоцитов и сниженным содержанием CD8⁺-клеток, что обусловило более высокие значения иммунорегуляторного индекса по сравнению с носителями генотипов GH/HaeIII AA и GDF9/AspEIGG. В ходе анализа продуктивных качеств выявлено преимущество генотипа CASTMM над CASTMN по показателям живой массы. Аналогичная закономерность установлена для генотипа GHAA в сравнении с GHAB. Полученные данные демонстрируют перспективность использования молекулярно-генетических маркеров в селекционных программах для одновременного улучшения продуктивных характеристик и укрепления иммунного статуса овец.

Ключевые слова: селекция маркеров, генотипирование, мелкий рогатый скот, иммунная реактивность, продуктивность

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Introduction/Введение

In sheep breeding, a key priority is the assessment and improvement of productivity, which depends on both genetic and environmental factors [1–5]. Breeding approaches that account for these factors are widely used in agricultural production and have contributed to better meat quality [6–9]. In practice, slaughter evaluation provides core indicators such as dressing percentage and carcass weight, which can correlate with specific genotypes [10–14]. Combining traditional methods with molecular tools, including genetic markers, enables systematic analysis of genetic resources and accelerates breeding programs¹ [15, 16]. Because many economically important traits are complex, results across breeds can be heterogeneous; continued study is needed to refine gene–trait links, manage genetic diversity, and improve prediction of productivity [17–21].

Maintaining diversity is also essential to avoid inbreeding depression under intensive selection schemes [22, 23]. For effective use of selection tools, the genetic structure of local populations must be described [24]. Identifying associations between gene variants and meat-production traits enables development of assays and, ultimately, genomic selection pipelines [25–28].

The Dagestan Mountain (DM) sheep and several other regionalized breeds remain understudied in this respect [29–31]. Marker-assisted selection (MAS) in sheep increasingly relies on well-characterized candidate genes. Polymorphisms in GH, CAST (calpastatin), and GDF9 (growth differentiation factor 9) have been extensively studied across breeds, with reported associations to growth, carcass/tenderness, and reproductive traits (GH [31–34], CAST [35–37], GDF9 [38–40]).

Building on this evidence, our study quantifies allele/genotype frequencies in [population] and tests genotype–phenotype associations relevant for selection. Improving economically important traits in DM sheep while conserving breed specificity is expected to raise meat quality and sustain adaptability to harsh local environments. Introducing validated DNA markers into selection schemes for DM sheep and relevant crossbreeds can increase selection efficiency and positively impact production metrics [41, 42].

Despite extensive evidence for GH, CAST and GDF9, data for Dagestan Mountain sheep and regional crosses are lacking. Given the breed's economic role and the need for rapid on-farm decisions, locally validated marker–trait associations are required to de-risk selection and improve prediction of growth and carcass outcomes. Therefore, we aimed to validate

polymorphisms in GH, CAST, and GDF9 in DM sheep and crosses, quantify their associations with growth, carcass, and immune traits, and derive selection-ready recommendations for marker-assisted decisions under farm conditions.

Materials and methods /

Материалы и методы исследования

The study was conducted at Agrofirma “Sogratl”, Gunibsky District, Republic of Dagestan, Russia, during 2022–2023 under lowland and mountainous grazing. The cohort comprised 114 crossbred sheep (Dagestan Mountain × Russian Meat Merino; 85 ewes, 29 rams) maintained under comparable feeding and housing.

Observational design to evaluate genotype–phenotype associations relevant for selection. Primary factor (fixed effect): genotype at candidate loci (GH, CAST, GDF9); levels used in analyses: GH (AA, AB), CAST (MM, MN), GDF9 (AA, AG, GG). Covariates: sex; age or measurement stage (where applicable).

Primary outcomes: longitudinal live weight; carcass traits (slaughter subset); immune cell subsets with immune-regulatory index (IRI = CD4⁺/CD8⁺). Reporting of genetic associations aligns with STREGA. Jugular venous blood (5–10 ml) was collected aseptically into heparinized vacutainers (25 U/ml). Samples were stored at 4 °C for up to 24 h or frozen at –20 °C until DNA extraction. Genomic DNA was isolated with DIALOM™ DNAPrep (IsoGeneLab, Moscow); typical yield 3–5 µg/100 µl, A260/280 = 1.6–2.0. Three loci were genotyped by PCR-RFLP: GH (HaeIII), CAST (MspI), GDF9 (BstHII).

Primer pairs/annealing temperatures and expected amplicons are summarized in Table 1. PCRs were performed in 20–25 µl reactions (1 × buffer, 1.5–2.0 mM MgCl₂, 0.2 mM each dNTP, 0.4 µM each primer, 1 U Taq polymerase) with cycling 95 °C 3 min; 35 cycles of 95 °C 30 s, annealing 60–63 °C 30 s, 72 °C 30 s; final 72 °C 5 min. Amplicons were digested with the specified endonucleases and resolved on 2–3% agarose; two independent readers called genotypes from fragment patterns [34].

The samples were stored at 4 °C for up to 24 hours or frozen at –20 °C until DNA extraction for PCR-RFLP genotyping of GH, GDF9, and CAST genes Table 1.

The experiment was conducted in compliance with the requirements set out in Directive of the European Parliament and of the Council of the European Union of 22 September 2010 No. 2010/63/EU on the protection of animals used for scientific purposes and the principles² of treatment of animals in accordance with Article 4 of Federal Law No. 498-FZ³.

¹ Оценка изменчивости генов, ассоциированных с живой массой и шерстной продуктивностью овец: научно-методические рекомендации / Г.М. Гончаренко, Т.Н. Хашируев, С.М. Даширмаев и др. Чита: Институт развития образования Забайкальского края. 2024; 35. ISBN 978-5-6045472-6-7, EDN WJGWSL

² Directive of the European Parliament and of the Council of the European Union On the protection of animals used for scientific purposes. https://ruslasa.ru/wp-content/uploads/2017/06/Directive_201063_rus.pdf

³ Федеральный закон от 20.12.2018 № 498-ФЗ (ред. от 24.07.2023) «Об ответственном обращении с животными и о внесении изменений в отдельные законодательные акты Российской Федерации».

Table 1. Primers and PCR-RFLP conditions for candidate loci

Gene	Nucleotide sequences	T (°C), Annealing	Amplicon, bp	Endonuclease	Genotypes
CAST	F:5'-tggggccaatgacgccatcgatg-3' R:5'-gggtggagcactctgatcacc-3'	62	422	MspI	MM/MN
GH	F:5'-ggaggcaggaaggatgaa-3' R:5'-ccaaggaggaggagagacaga-3'	60	277	HaeIII	AA/AB/BB
GDF9	F:5'-gaagactggatgggaaatg-3' R:5'-ccaatctgctctacacac-3'	63	462	BstHII	AA/AG/GG

Cellular immunity was assessed in carriers of selected GH and GDF9 genotypes by flow cytometry. Whole blood (EDTA) was stained with fluorochrome-conjugated monoclonal antibodies to CD3, CD4, CD8, and CD21 with appropriate isotype controls; erythrocytes were lysed with a commercial kit. Data were acquired on an Attune™ NxT Acoustic Focusing Flow Cytometer (Thermo Fisher Scientific, USA), a benchtop analyzer that uses acoustic-focusing fluidics; the system supports configurations with four lasers (405/488/561/637 nm) and up to 14 colors, and runs Attune Cytometric Software for acquisition and analysis.

Individual live weight was recorded at predefined stages birth, pre-transhumance, arrival on mountain pastures, weaning, pre-winter, arrival on winter pastures) using low-profile platform livestock scales Promves-PL-VZh (Promvesoborudovanie LLC, Yekaterinburg, Russia) — a non-automatic weighing instrument, OIML R 76 Class III, configured for static weighing with strain-gauge load cells. The model series is listed in the Russian State Register of Measuring Instruments (entry No. 87205-22; valid until 24 Oct 2027; inter-verification interval 1 year).

The farm configuration provided capacity 600–2000 kg (series options up to 40 000 kg), graduation 0.1–0.2 kg, and a 1000 × 2000 mm platform. Before each weighing session, the scale was checked with certified standard weights; two consecutive readings were averaged (difference ≤ 0.2 kg). Measurements adhered to the principles for non-automatic weighing instruments in OIML R 76; for national practice we also reference GOST 25955-83⁴ (methods for determining sheep productivity parameters). All animals were clinically healthy, and the conditions of their maintenance complied with zootechnical standards and zoohygienic requirements⁵.

Analyses were performed in R (v4.x) using lme4 for mixed models and emmeans for estimated marginal means; HardyWeinberg was used for Hardy — Weinberg tests. Live

weight (repeated measures) was analysed with a linear mixed model weight — genotype + stage + genotype: stage + sex + (1|animal); we report estimated marginal means (EMMs) with Tukey multiplicity adjustment for within-factor contrasts, effects (β), 95% CIs and two-sided P. Immunology traits were modelled as trait — genotype + sex + age + (1|batch); normality and homoscedasticity were checked on residuals, with log-transform or a GLMM alternative if assumptions were violated; EMMs are reported with multiplicity control.

Carcass traits were analysed with linear (or mixed) models trait — genotype + sex; given the small slaughter subset, we emphasised effect sizes with 95% CIs and used robust / permutation-based checks if normality was untenable. For genotypes, allele/genotype frequencies were computed and HWE was assessed per locus using the exact test (Hardy — Weinberg — HWExact) with a χ^2 sensitivity check; rare, unobserved levels (e. g., CAST NN) are explicitly documented. Across families of outcomes we additionally controlled the false-discovery rate (BH-FDR, $q < 0.05$). Factor coding and reference levels are specified in the Results/Supplement. Reporting of design, sample size, and statistics follows ARRIVE 2.0 and STREGA recommendations.

Results and discussion /

Результаты и обсуждение

The importance of the immune system for growth and development is especially relevant during the growth period of lambs, since it is during this period that the immune status is established.

The study of the levels of T — and B-lymphocytes in the blood of lambs of different genotypes made it possible to assess their immune potential Table 2.

It was found, that the Dagestan Mountain lambs with the genotypes GH/HaeIII AB and GDF9/AspIEIAA had a higher level of T-helpers and lower T-suppressors,

Table 2. Indicators of immune reactivity of lambs of different genotypes

Genotypes by genes	Indicator				
	Immune reactivity, 10 ⁹ /l				IRI
	T-cells	B-cells	T-suppressors	T-helpers	
Dagestan mountain breed of sheep (n = 30)					
GH ^{AA}	0.60 ± 0.09	0.52 ± 0.06	0.34 ± 0.04	0.23 ± 0.16	0.68
GH ^{AB}	0.69 ± 0.12	0.53 ± 0.08	0.26 ± 0.15	0.29 ± 0.06	1.1
GDF9 ^{GG}	0.59 ± 0.08	0.35 ± 0.1	0.40 ± 0.14	0.28 ± 0.14	0.7
GDF9 ^{AA}	0.66 ± 0.15	0.40 ± 0.14	0.3 ± 0.1	0.35 ± 0.16	1.2
GDF9 ^{AG}	0.61 ± 0.08	0.35 ± 0.12	0.38 ± 0.12	0.26 ± 0.14	0.68
Crossbred lambs (n = 20)					
GH/HaeIII ^{AA}	0.57 ± 0.08	0.42 ± 0.06	0.36 ± 0.14	0.37 ± 0.05	1.03
GH/HaeIII ^{AB}	0.62 ± 0.14	0.50 ± 0.12	0.42 ± 0.08	0.38 ± 0.14	0.9
GDF9/AspI ^{GG}	0.5 ± 0.06	0.42 ± 0.16	0.3 ± 0.04	0.37 ± 0.04	1.23
GDF9/AspI ^{AA}	0.65 ± 0.12	0.47 ± 0.16	0.32 ± 0.06	0.3 ± 0.02	0.94
GDF9/AspI ^{AG}	0.54 ± 0.14	0.46 ± 0.06	0.34 ± 0.06	0.35 ± 0.02	1.03

⁴ ГОСТ 25955-83 Сельскохозяйственная племенная продуктивность. Методы определения показателей продуктивности овец. М.: Издательство стандартов. 1983; 13.

⁵ ГОСТ 34088-2017 Руководство по размещению и уходу за лабораторными животными. Правила содержания и ухода за сельскохозяйственными животными. М.: Межгосударственный совет по стандартизации, метрологии и сертификации. 2018; 36.

which affected on the immunoregulatory index (IRI), making it higher, compared to the lambs of the genotypes GH/HaeIII AA and GDF9/AspIcGG.

Crossbred lambs also had a higher number of B — and T-lymphocytes with genotypes GH/HaeIII AB and GDF9/AspIcIAA. The studies show, that the body's reactivity on negative environmental influences is specific for each genotype and is apparently determined by an innate genetic program. This program allows to increase production of T — and B-lymphocytes, as well as T-helpers, but decrease the number of T-suppressors. Such regulation helps to correct and strengthen of the adaptation and compensation systems in the body on various stages of its development.

With the development of molecular genetics, new opportunities are opening up for optimizing of breeding selection and increasing of the efficiency of livestock farms.

A key aspect is the identification of genotypes, that can improve the productive characteristics of animals. Particular attention is paid to genes, that affect on productive qualities, including the growth hormone gene (GH) and calpain (CAST).

In the context of studying of the effect of genetic markers on the weight of Dagestan Mountain lambs, their weight was measured on various stages of life.

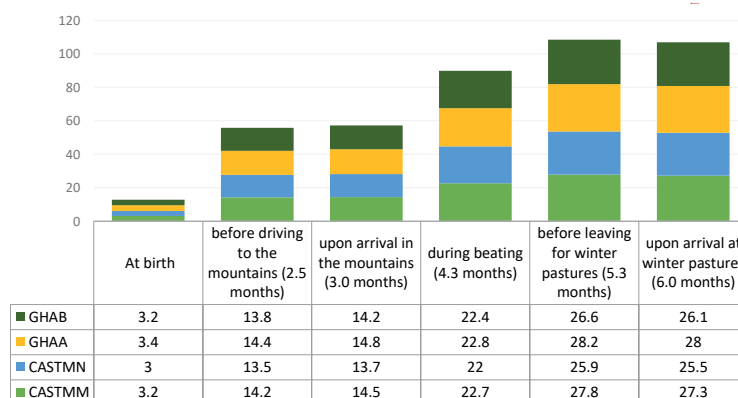
This made it possible to evaluate the relationship between various forms of the CAST and GH genes and changes of live weight (Fig. 1).

Analysis of the sheep weight showed, that the greatest increase of weight was observed before the period of grazing on summer pastures, when the average gain was 14.6 kg. Also, a significant gain 23.0 kg was recorded in the time of separation of mothers from lambs. At the same time, the lowest weight indicators were noted during the period of movement between winter and summer pastures, while their average weight was 27.1 kg.

During the study of various stages of development of experimental individuals, it was revealed, that the highest rates of weight gain were observed in individuals with certain genetic markers. Thus, individuals, carrying a combination of MM on the CAST gene and AA on the GH gene, demonstrated the best results in body weight gain and daily weight gain. It is noted, that the sample of analysis did not include individuals with the CASTNN and GHBB genotypes, which excludes the possibility of their assessment within the framework of the study. The study showed, that animals with the CASTMM genotype have a larger mass on average, compared to their relatives with the CASTMN genotype.

Observations in different periods revealed the following differences in mass: before being taken to pasture, the difference was 4.9%, and when the cattle reached the mountain pastures — 5.5%. After the summer period, during weaning, the gap decreased

Fig. 1. Live weight indicators of Dagestan Mountain ewe lambs of different genotypes



to 3.1%. However, before being sent to wintering and upon arrival there, the difference increased to 6.9 and 6.6% respectively.

In relation to the GH gene, differences in weight gain were also observed between carriers of the GHAA and GHAB genotypes.

Thus, when grazing on summer pastures, animals with the GHAA genotype had a 4.2% weight advantage. When the cattle reached summer pastures, the weight difference was — 4.8%. The difference between carriers of the AA and AB alleles increased before being sent to wintering and upon arrival there, and amounted to 5.7 and 6.8%, respectively.

The study showed, that certain genotypes, namely GHAA and CASTMM, demonstrate better results in live weight and weight gain, compared to GHAB and CASTMN. The observed differences in productivity become especially noticeable during key periods of cattle breeding, such as the transfer of animals to summer and winter pastures. At critical moments, such as before the cattle are driven to the mountains and back, as well as during of the change of seasonal pastures, the advantage of the GHAA and CASTMM genotypes becomes obvious.

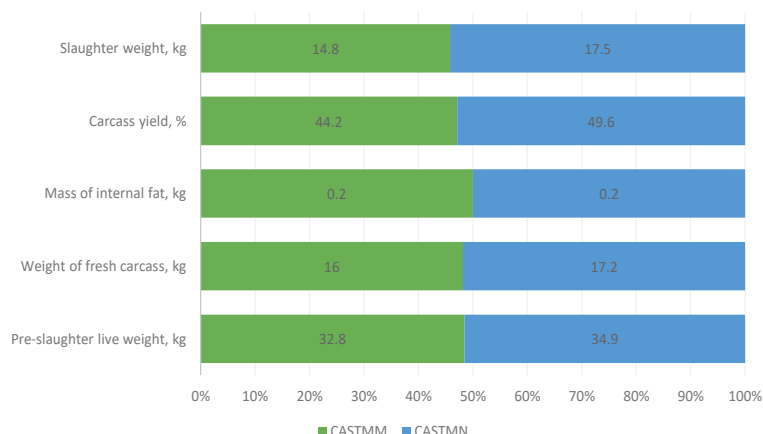
To analyze the quality characteristics of meat of different sheep genotypes, including its composition and nutritional value, control slaughters are carried out. During the study, crossbred rams with the CASTMM and CASTMN genotypes were weighed individually. This allowed us to select animals with similar weights for slaughter. Before the slaughter procedure, specialists examined the sheep to ensure their health and readiness for slaughter.

The study of different genotypes of CASTMM and CASTMN sheep showed interesting differences in their parameters before and after slaughter.

Animals with the CASTMM genotype (Fig. 2) demonstrated better results on number of indicators.

When evaluating of the carcass after slaughter, the difference was also noticeable: CASTMM ewes surpassed CASTMN on 7.0% in terms of fresh carcass weight. While in terms of internal fat content, ewes with the CASTMN and CASTMN genotypes showed equal values. The percentage of carcass weight to live weight was on 10.9% higher in CASTMM, indicating

Fig. 2. Results of control slaughter of crossbred rams with genotypes CASTMM and CASTMN



on their more favorable slaughter qualities. In general, compared to CASTMN, the slaughter weight in ewes with the CASTMM genotype was almost on 15.4% higher, which can play a significant role in commercial breeding.

Все авторы несут ответственность за работу и представленные данные. Все авторы внесли равный вклад в работу. Авторы в равной степени принимали участие в написании рукописи и несут равную ответственность за плагиат. Авторы объявили об отсутствии конфликта интересов.

Conclusions/Выводы

Based on the study results, the CASTMM genotype demonstrates statistically significant advantages for carcass-related traits under the evaluated farm conditions. The GHAA genotype shows a consistent association with pre-weaning growth performance in lambs.

The confirmed marker-trait relationships support the practical application of GH and CAST markers in marker-assisted selection programs for Dagestan Mountain sheep and their crossbred populations.

To enhance the generalizability and reliability of these estimates, further validation across larger and more diverse multi-farm cohorts is recommended. These findings provide valuable insights for breeding programs aimed at improving meat productivity traits in sheep populations.

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REFERENCES

- Rani C. *et al.* Genetic variation in host immune response to major infectious diseases in bovines and its application in animal breeding: a review. *Exploratory Animal and Medical Research*. 2023; 13(2): 146–157. <https://doi.org/10.52635/eamr/13.2.146-157>
- Kosimov M.A., Yuldashbaev Yu.A., Kosimov A.M., Pogosyan G.A., Mamedov A.A. *ogl.* Trends in the development of sheep breeding in the world. *Sheep, goats, wool business*. 2025; (1): 3–5 (in Russian). <https://doi.org/10.26897/2074-0840-2025-1-3-5>
- Chernobay E.N., Rezun N.A., Onishchenko O.N., Ismailov I.S. Current state and methods of increasing sheep productivity. *Genomics and biotechnology in agriculture: a collection of scientific articles based on the materials of the plenary session of the 88th scientific and practical conference of the Stavropol State Agricultural University "Agrarian Science – to the North Caucasus Federal District"*. Stavropol: Stavropol State Agrarian University. 2023; 48–52 (in Russian). <https://elibrary.ru/crjwgj>
- Samusenko L.D., Mamaev A.V., Khimicheva S.N., Rodina N.D., Sergeeva E.Yu. A model of bioenergetic assessment and forecasting of wool productivity in sheep breeding. *Agrarian science*. 2025; (9): 54–61 (in Russian). <https://doi.org/10.32634/0869-8155-2025-398-09-54-61>
- Yuldashbaev Yu.A., Korneenko-Zhilyaeva S.A., Pakhomova E.V., Naumenko I.B. Prospects for the development of meat and dairy sheep breeding. *Zootechnical and veterinary science – the basis for the innovative development of animal husbandry in Russia. Collection of articles, based on the materials of the All-Russian scientific and practical conference with international participation, dedicated to the 90th anniversary of the founding of the Institute of Animal Science and Biology of the Russian State Agrarian University – Moscow Agricultural Academy named after K.A. Timiryazev*. Moscow: Russian State Agrarian University – Moscow Timiryazev Agricultural Academy. 2024; 128–133 (in Russian). <https://elibrary.ru/nqqphm>
- Eusebi P.G., Martinez A., Cortes O. Genomic Tools for Effective Conservation of Livestock Breed Diversity. *Diversity*. 2019; 12(1): 8. <https://doi.org/10.3390/d12010008>

БИБЛИОГРАФИЧЕСКИЙ СПИСОК

- Rani C. *et al.* Genetic variation in host immune response to major infectious diseases in bovines and its application in animal breeding: a review. *Exploratory Animal and Medical Research*. 2023; 13(2): 146–157. <https://doi.org/10.52635/eamr/13.2.146-157>
- Косимов М.А., Юлдашбаев Ю.А., Косимов А.М., Погосян Г.А., Мамедов А.А. *оглы.* Тенденции развития овцеводства в мире. *Овцы, козы, шерстяное дело*. 2025; (1): 3–5. <https://doi.org/10.26897/2074-0840-2025-1-3-5>
- Чернобай Е.Н., Резун Н.А., Онищенко О.Н., Исмаилов И.С. Современное состояние и методы повышения продуктивности овец. *Геномика и биотехнологии в сельском хозяйстве: сборник научных статей по материалам пленарного заседания 88-й научно-практической конференции ФГБОУ ВО «Ставропольский ГАУ» «Аграрная наука – Северо-Кавказскому федеральному округу»*. Ставрополь: Ставропольский государственный аграрный университет. 2023; 48–52. <https://elibrary.ru/crjwgj>
- Самусенко Л.Д., Мамаев А.В., Химичева С.Н., Родина Н.Д., Сергеева Е.Ю. Модель биоэнергетической оценки и прогнозирования шерстной продуктивности в овцеводстве. *Аграрная наука*. 2025; (9): 54–61. <https://doi.org/10.32634/0869-8155-2025-398-09-54-61>
- Юлдашбаев Ю.А., Корнеенко-Жильева С.А., Пахомова Е.В., Науменко И.Б. Перспективы развития мясного и молочного овцеводства. *Зоотехническая и ветеринарная наука – основа инновационного развития животноводства России. Сборник статей по материалам Всероссийской научно-практической конференции с международным участием, посвященной 90-летию со дня образования Института зоотехнии и биологии РГАУ – МСХА им. К.А. Тимирязева*. М.: Российский государственный аграрный университет – МСХА им. К.А. Тимирязева. 2024; 128–133. <https://elibrary.ru/nqqphm>
- Eusebi P.G., Martinez A., Cortes O. Genomic Tools for Effective Conservation of Livestock Breed Diversity. *Diversity*. 2019; 12(1): 8. <https://doi.org/10.3390/d12010008>

7. Galanova E.S., Bobryshov S.S., Karpova E.D. Current trends and prospects for the development of sheep breeding productivity by crossing domestically selected ewes with imported rams. *Promising developments of young scientists in the field of veterinary medicine, production and processing of agricultural products*. Stavropol: Stavropol State Agrarian University. 2024; 103–108 (in Russian). <https://elibrary.ru/tdvohv>
8. Golaev Sh.Kh., Galanova E.S., Bondar A.V., Filipenkova V.A. Prospects for linear breeding in sheep breeding. *Promising developments of young scientists in the field of veterinary medicine, production and processing of agricultural products*. Stavropol: Stavropol State Agrarian University. 2024; 48–51 (in Russian). <https://elibrary.ru/bcssvx>
9. Abdurasulov A.Kh., Kalandarov M.A., Rebezov M.B., Kosilov V.I. Live weight and body features of Ala-Too sheep. *Agrarian science*. 2025; (10): 71–80 (in Russian). <https://doi.org/10.32634/0869-8155-2025-399-10-71-80>
10. Baranova N.S., Furaeva N.S., Zvereva E.A., Korolev A.A., Vorobyeva S.S. Determination of optimal blood parameters for the Brown Swiss breed when improving Kostroma breed cattle. *Dairy and Beef Cattle Farming*. 2024; (4): 13–18 (in Russian). <https://doi.org/10.33943/MMS.2024.51.19.003>
11. Andrienko D.A., Traisov B.B., Rebezov M.B., Galieva Z.A. Morphological composition of carcasses of young Stavropol sheep in the Southern Urals. *Science and Education*. 2020; 3–1(60): 20–25 (in Russian). <https://elibrary.ru/gpvhdh>
12. Dnekeshev A.K., Musa R.K. Ratio of hot carcass and by-products to live weight at different ages of castration of rams. *Modern problems and technologies in animal husbandry. Collection of works based on the results of the All-Russian (national) scientific and practical conference dedicated to the 95th anniversary of the birth of the Honored Scientist of the Russian Federation, Honorary Worker of the Higher Professional Education of the Russian Federation, Head of the Department of Breeding Farm Animals (from 1981 to 1991), Doctor of Agricultural Sciences, Professor of the Nizhny Novgorod State Agricultural Academy L.P. Prakhov*. Nizhny Novgorod: Nizhny Novgorod State Agrotechnological University named after L.Ya. Florentyev. 2024; 112–118 (in Russian). <https://elibrary.ru/xheltc>
13. Belookova O.V., Rebezov M.B., Lennikova M.G., Zybilitseva M.A. Influence of zoohygienic parameters on meat productivity of pigs. *Bulletin of Osh State University. Agriculture: agronomy, veterinary science and animal husbandry*. 2024; 4(9): 52–64 (in Russian). [https://doi.org/10.52754/16948696_2024_4\(9\)_8](https://doi.org/10.52754/16948696_2024_4(9)_8)
14. Gritsenko S.A., Belookov A.A., Ermolov S.M., Rebezov M.B., Gritsenko M.D. Exterior indicators of maternal breed pigs at different stages of ontogenesis. *Agrarian science*. 2025; (9): 45–53 (in Russian). <https://doi.org/10.32634/0869-8155-2025-398-09-45-53>
15. Sahib M.R., AL-Jaryan I.L., Al-Thuwaini T.M. GDF9 gene c. 260G > A mutation and sheep litter size: a meta-analysis. *Journal of the Hellenic Veterinary Medical Society*. 2024; 75(2): 7493–7500. <https://doi.org/10.12681/jhvms.35128>
16. Guseinova Z.M. New candidate genes associated with meat productivity traits in sheep. *Current problems and prospects for the development of the agro-industrial complex of the Republic of Dagestan. Proceedings of the regional scientific and practical conference*. Makhachkala: Individual entrepreneur S.A. Magomedaliyev. 2024; 70–74 (in Russian). <https://elibrary.ru/tlvhgm>
17. Krivoruchko A.Yu., Skokova A.V., Yatsyk O.A., Kanibolotskaya A.A. Modern approaches to the genetic identification of farm animal breeds (review). *Agricultural Science Euro-North-East*. 2021; 22(3): 317–328 (in Russian). <https://doi.org/10.30766/2072-9081.2021.22.3.317-328>
18. Alieva P.O. A new approach to the selection of candidate genes in sheep breeding. *Actual problems and prospects for the development of the agro-industrial complex of the Republic of Dagestan: Proceedings of the regional scientific and practical conference*. Makhachkala: Individual entrepreneur S.A. Magomedaliyev. 2024; 34–39 (in Russian). <https://elibrary.ru/pnaswe>
19. Nikonova E.A. et al. Efficiency of growing purebred and crossbred rams. *Journal of Osh State University. Agriculture: agronomy, veterinary and zootechnics*. 2023; (4): 164–170 (in Russian). https://doi.org/10.52754/16948696_2023_4_24
7. Галанова Е.С., Бобрышов С.С., Карпова Е.Д. Современные тенденции и перспективы развития продуктивности овцеводства при скрещивании маток отечественной селекции с баранами импортных пород. *Перспективные разработки молодых ученых в области ветеринарии, производства и переработки сельскохозяйственной продукции*. Ставрополь: Ставропольский государственный аграрный университет. 2024; 103–108. <https://elibrary.ru/tdvohv>
8. Голаев Ш.Х., Галанова Е.С., Бондарь А.В., Филипенкова В.А. Перспективы линейного разведения в овцеводстве. *Перспективные разработки молодых ученых в области ветеринарии, производства и переработки сельскохозяйственной продукции*. Ставрополь: Ставропольский государственный аграрный университет. 2024; 48–51. <https://elibrary.ru/bcssvx>
9. Абдурасулов А.Х., Каландаров М.А., Ребезов М.Б., Косилов В.И. Живая масса и особенности телосложения овец породы ала-тоо. *Аграрная наука*. 2025; (10): 71–80. <https://doi.org/10.32634/0869-8155-2025-399-10-71-80>
10. Баранова Н.С., Фураева Н.С., Зверева Е.А., Королев А.А., Воробьева С.С. Определение оптимальных параметров кровности по бурой швицкой породе при совершенствовании скота костромской породы. *Молочное и мясное скотоводство*. 2024; (4): 13–18. <https://doi.org/10.33943/MMS.2024.51.19.003>
11. Андриенко Д.А., Траисов Б.Б., Ребезов М.Б., Галиева З.А. Морфологический состав туш молодняка овец ставропольской породы на Южном Урале. *Наука и образование*. 2020; 3–1(60): 20–25. <https://elibrary.ru/gpvhdh>
12. Днекешев А.К., Муса Р.К. Соотношение парной туши и субпродуктов к живой массе при разных возрастах кастрации баранчиков. *Современные проблемы и технологии в животноводстве. Сборник трудов по итогам Всероссийской (национальной) научно-практической конференции, посвященной 95-летию со дня рождения заслуженного деятеля науки РФ, почетного работника ВПО РФ, заведующего кафедрой разведения сельскохозяйственных животных (с 1981 по 1991 г.) д-ра с.-х. наук, профессора Нижегородской государственной сельскохозяйственной академии Л.П. Прахова*. Нижний Новгород: Нижегородский государственный агротехнологический университет им. Л.Я. Флорентьева. 2024; 112–118. <https://elibrary.ru/xheltc>
13. Белоокова О.В., Ребезов М.Б., Ленникова М.Г., Зяблицева М.А. Влияние зоогигиенических параметров на мясную продуктивность свиней. *Вестник Ошского государственного университета. Сельское хозяйство: агрономия, ветеринария и зоотехния*. 2024; 4(9): 52–64. [https://doi.org/10.52754/16948696_2024_4\(9\)_8](https://doi.org/10.52754/16948696_2024_4(9)_8)
14. Гриценко С.А., Белооков А.А., Ермолов С.М., Ребезов М.Б., Гриценко М.Д. Показатели экстерьера свинок материнской породы на различных этапах онтогенеза. *Аграрная наука*. 2025; (9): 45–53. <https://doi.org/10.32634/0869-8155-2025-398-09-45-53>
15. Sahib M.R., AL-Jaryan I.L., Al-Thuwaini T.M. GDF9 gene c. 260G > A mutation and sheep litter size: a meta-analysis. *Journal of the Hellenic Veterinary Medical Society*. 2024; 75(2): 7493–7500. <https://doi.org/10.12681/jhvms.35128>
16. Гусейнова З.М. Новые гены — кандидаты, связанные с признаками мясной продуктивности овец. *Актуальные проблемы и перспективы развития АПК Республики Дагестан. Материалы региональной научно-практической конференции*. Махачкала: С.А. Магомедалиев. 2024; 70–74. <https://elibrary.ru/tlvhgm>
17. Криворучко А.Ю., Скокова А.В., Яцык О.А., Каниболоцкая А.А. Современные подходы генетической идентификации породной принадлежности сельскохозяйственных животных (обзор). *Аграрная наука Евро-Северо-Востока*. 2021; 22(3): 317–328. <https://doi.org/10.30766/2072-9081.2021.22.3.317-328>
18. Алиева П.О. Новый подход к подбору генов-кандидатов в овцеводстве. *Актуальные проблемы и перспективы развития АПК Республики Дагестан. Материалы региональной научно-практической конференции*. Махачкала: С.А. Магомедалиев. 2024; 34–39. <https://elibrary.ru/pnaswe>
19. Никонова Е.А. и др. Эффективность выращивания чистопородных и помесных баранчиков. *Вестник Ошского государственного университета. Сельское хозяйство: агрономия, ветеринария и зоотехния*. 2023; (4): 164–170. https://doi.org/10.52754/16948696_2023_4_24

20. Gerasimova T.G., Rebezov M.B., Lukin E.V. Wool productivity of sheep of different genotypes. *Actual problems of intensive development of animal husbandry. Collection of papers based on the materials of the national scientific and practical conference with international participation, dedicated to the memory of the Honored Worker of Higher Education of the Russian Federation, Honorary Worker of Higher professional education of the Russian Federation, honorary citizen of the Bryansk region, Doctor of Biology, Professor E.P. Vashchekin*. Bryansk: Bryansk State Agrarian University. 2022; 300–303 (in Russian). <https://elibrary.ru/fkpdfx>
21. Kosilov V.I. *et al.* The effect of the genotype of young sheep on the nitrogen balance. *Agrarian science*. 2025; (4): 81–87 (in Russian). <https://doi.org/10.32634/0869-8155-2025-393-04-81-87>
22. Milchevsky V.D., Dvalishvili V.G., Zhiryakov A.M. On inbreeding and rotations in pair selection in sheep flocks. *Sheep, goats, wool business*. 2017; (2): 4–7 (in Russian). <https://elibrary.ru/yrrkuv>
23. Lakota E.A., Vorontsova O.A. Exterior-productive indicators of Stavropol breed sheep aged 13–14 months with inbreeding selection. *Agrarian science*. 2023; (8): 65–71 (in Russian). <https://doi.org/10.32634/0869-8155-2023-373-8-65-71>
24. Dzhatdov Kh.M., Erkenov T.A. Polymorphism of non-coding sequences in Karachai sheep. *Agrarian science*. 2024; (12): 109–112 (in Russian). <https://doi.org/10.32634/0869-8155-2024-389-12-109-112>
25. Daveteeva M.A. Modern methods of genotyping in animal husbandry. *A modern view on the development of the fishing industry complex. Proceedings of the All-Russian scientific and practical conference (within the framework of the "Priority 2030" program)*. Makhachkala: Dagestan State Agrarian University named after M.M. Dzhambulatov. 2024; 286–297 (in Russian). <https://elibrary.ru/iwyrso>
26. Zabelina M.V., Amiyana A.A. Evaluation of the slaughter indicators of the Edilbaevsky sheep breed depending on the milk productivity of sheep. *Agrarian science*. 2024; (8): 112–117 (in Russian). <https://doi.org/10.32634/0869-8155-2024-385-8-112-117>
27. Moroz V.A., Chernobai E.N., Novgorodova N.A., Serdyukov I.G. Live weight and exterior characteristics of sheep from homogeneous and heterogeneous selection. *Bulletin of the Kurgan State Agricultural Academy*. 2017; 2(22): 51–53 (in Russian).
28. Lakota E.A. The influence of parental age on the productive qualities of offspring sheep. *Agrarian science*. 2025; (7): 57–61 (in Russian). <https://doi.org/10.32634/0869-8155-2025-396-07-57-61>
29. Zinovieva N.A. *et al.* Genome-wide SNP analysis clearly distinguished the Belarusian Red cattle from other European cattle breeds. *Animal Genetics*. 2021; 52(5): 720–724. <https://doi.org/10.1111/age.13102>
30. Petherbridge G. *et al.* Verkhnee Gakvari: The contribution of adat, religious beliefs and public education to collective environmental management in an agro-pastoral community in the Dagestan high Caucasus. *South of Russia: ecology, development*. 2021; 16(3): 142–179. <https://doi.org/10.18470/1992-1098-2021-3-142-179>
31. Prozherin V.P., Yaluga V.L. Holmogorskaya breed. *Dairy and Beef Cattle Farming*. 2020; (7): 10 (in Russian). <https://elibrary.ru/brmwde>
32. Pelmuş R.Ş., Grosu H., Rotar M.C., Gras M.A., Lazăr C., Popa F. The Genetic Influence on Sheep Meat Quality, Growth and Body Weight: A Review. *Asian Journal of Dairy and Food Research*. 2020; 39(3): 225–231. <https://doi.org/10.18805/ajdfr.DR-176>
33. El-Mansy S.A. *et al.* The growth hormone gene polymorphism and its relationship to performance and carcass features in Egyptian Awassi lambs. *Heliyon*. 2023; 9(3): e14194. <https://doi.org/10.1016/j.heliyon.2023.e14194>
34. Tanış A.N., Keskin İ. Association between growth hormone gene polymorphism and some growth traits in Akkaraman and Anatolian Merino sheep. *Molecular Biology Reports*. 2024; 52: 54. <https://doi.org/10.1007/s11033-024-10126-2>
35. Dagdelen U., Esenbuga N. Association of GH/HaeIII polymorphism with growth and developmental traits of Morkaraman sheep. *Archives Animal Breeding*. 2025; 68(3): 575–587. <https://doi.org/10.5194/aab-68-575-2025>
36. Greguła-Kania M., Gruszecki T.M., Junkuszew A., Juszcuk-Kubiak E., Florek M. Association of CAST gene polymorphism with carcass value and meat quality in two synthetic lines of sheep. *Meat Science*. 2019; 154: 69–74. <https://doi.org/10.1016/j.meatsci.2019.04.007>
37. Jawasreh K.I., Al-Amareen A.H., Aad P.Y. Relationships between HhaI Calpastatin Gene Polymorphism, Growth Performance, and Meat Characteristics of Awassi Sheep. *Animals*. 2019; 9(9): 667. <https://doi.org/10.3390/ani9090667>
20. Герасимова Т.Г., Ребезов М.Б., Лукин Е.В. Шерстная продуктивность овец разного генотипа. *Актуальные проблемы интенсивного развития животноводства. Сборник трудов по материалам национальной научно-практической конференции с международным участием, посвященной памяти заслуженного работника Высшей школы РФ, почетного работника высшего профессионального образования РФ, почетного гражданина Брянской области д-ра биол. наук, профессора Е.П. Ващекина*. Брянск: Брянский государственный аграрный университет. 2022; 300–303. <https://elibrary.ru/fkpdfx>
21. Косилов В.И. и др. Влияние генотипа молодняка овец на баланс азота. *Аграрная наука*. 2025; (4): 81–87. <https://doi.org/10.32634/0869-8155-2025-393-04-81-87>
22. Мильчевский В.Д., Двалишвили В.Г., Жиряков А.М. Об инбридинге и ротациях при подборе пар в стадах овец. *Овцы, козы, шерстяное дело*. 2017; (2): 4–7. <https://elibrary.ru/yrrkuv>
23. Лакота Е.А., Воронцова О.А. Экстерьерно-продуктивные показатели овец ставропольской породы в возрасте 13–14 месяцев при внутрипородном отборе. *Аграрная наука*. 2023; (8): 65–71. <https://doi.org/10.32634/0869-8155-2023-373-8-65-71>
24. Джатдоев Х.М., Эркенов Т.А. Полиморфизм некодирующих последовательностей у овец карачаевской породы. *Аграрная наука*. 2024; (12): 109–112. <https://doi.org/10.32634/0869-8155-2024-389-12-109-112>
25. Даветеева М.А. Современные методы генотипирования в животноводстве. *Современный взгляд на развитие рыбопромышленного комплекса. Материалы Всероссийской научно-практической конференции (в рамках программы «Приоритет-2030»)*. Махачкала: Дагестанский государственный аграрный университет им. М.М. Джамбулатова. 2024; 286–297. <https://elibrary.ru/iwyrso>
26. Забелина М.В., Амиян А.А. Оценка показателей убоя баранчиков эдилбаевской породы в зависимости от молочной продуктивности овцематок. *Аграрная наука*. 2024; (8): 112–117. <https://doi.org/10.32634/0869-8155-2024-385-8-112-117>
27. Мороз В.А., Чернобай Е.Н., Новгородова Н.А., Сердюков И.Г. Живая масса и экстерьерные особенности овец от однородного и разнородного подбора. *Вестник Курганской ГСХА*. 2017; 2(22): 51–53.
28. Лакота Е.А. Влияние возраста родителей на продуктивные качества потомства у овец. *Аграрная наука*. 2025; (7): 57–61. <https://doi.org/10.32634/0869-8155-2025-396-07-57-61>
29. Zinovieva N.A. *et al.* Genome-wide SNP analysis clearly distinguished the Belarusian Red cattle from other European cattle breeds. *Animal Genetics*. 2021; 52(5): 720–724. <https://doi.org/10.1111/age.13102>
30. Петербридж Г. и др. Верхнее Гаквари: вклад адата, религиозных убеждений и народного образования в коллективное управление окружающей средой в агропастырском сообществе Дагестанской части Большого Кавказа. *Юг России: экология, развитие*. 2021; 16(3): 142–179 (на англ. яз.). <https://doi.org/10.18470/1992-1098-2021-3-142-179>
31. Прожерин В.П., Ялуга В.Л. Холмогорская порода. *Молочное и мясное скотоводство*. 2020; (7): 10. <https://elibrary.ru/brmwde>
32. Pelmuş R.Ş., Grosu H., Rotar M.C., Gras M.A., Lazăr C., Popa F. The Genetic Influence on Sheep Meat Quality, Growth and Body Weight: A Review. *Asian Journal of Dairy and Food Research*. 2020; 39(3): 225–231. <https://doi.org/10.18805/ajdfr.DR-176>
33. El-Mansy S.A. *et al.* The growth hormone gene polymorphism and its relationship to performance and carcass features in Egyptian Awassi lambs. *Heliyon*. 2023; 9(3): e14194. <https://doi.org/10.1016/j.heliyon.2023.e14194>
34. Tanış A.N., Keskin İ. Association between growth hormone gene polymorphism and some growth traits in Akkaraman and Anatolian Merino sheep. *Molecular Biology Reports*. 2024; 52: 54. <https://doi.org/10.1007/s11033-024-10126-2>
35. Dagdelen U., Esenbuga N. Association of GH/HaeIII polymorphism with growth and developmental traits of Morkaraman sheep. *Archives Animal Breeding*. 2025; 68(3): 575–587. <https://doi.org/10.5194/aab-68-575-2025>
36. Greguła-Kania M., Gruszecki T.M., Junkuszew A., Juszcuk-Kubiak E., Florek M. Association of CAST gene polymorphism with carcass value and meat quality in two synthetic lines of sheep. *Meat Science*. 2019; 154: 69–74. <https://doi.org/10.1016/j.meatsci.2019.04.007>
37. Jawasreh K.I., Al-Amareen A.H., Aad P.Y. Relationships between HhaI Calpastatin Gene Polymorphism, Growth Performance, and Meat Characteristics of Awassi Sheep. *Animals*. 2019; 9(9): 667. <https://doi.org/10.3390/ani9090667>

38. Daldaban F., Karakus F., Arslan K., Cinar M.U., Akyuz B. The Role of *DGAT1-Alul* and *CAST-MspI* Polymorphisms in Early Growth Traits of Morkaraman Lambs. *Veterinary Medicine and Science*. 2025; 11(3): e70395. <https://doi.org/10.1002/vms3.70395>
39. Zhang Y. *et al.* Association of *BMP15* and *GDF9* Gene Polymorphisms with Litter Size in Hu Sheep. *Genes*. 2025; 16(2): 168. <https://doi.org/10.3390/genes16020168>
40. Budiyo A., Hartanto S., Widayanti R., Setyawan E.M.N., Wardono H.P., Gustari S. The relationship between G1 (c. 260 G > A) and G4 (c. 721 G > A) polymorphisms in the *GDF9* gene and the litter size of sheep: A meta-analysis study. *Journal of Advanced Veterinary and Animal Research*. 2023; 10(4): 599–607. <http://doi.org/10.5455/javar.2023.j715>
41. Ozdemirov A.A., Chizhova L.N., Khozhokov A.A., Surzhikova E.S., Dogeev G.D., Abdulmagomedov S.Sh. Polymorphism of genes *CAST*, *GH*, *GDF9* of sheep of the Dagestan mountain breed. *South of Russia: ecology, development*. 2021 16(2): 39–44 (in Russian). <https://doi.org/10.18470/1992-1098-2021-2-39-44>
42. Volkova V.V. *et al.* Investigation of the Genetic Diversity of Dagestan Mountain Cattle Using STR-Markers. *Diversity*. 2022; 14(7): 569. <https://doi.org/10.3390/d14070569>

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38. Daldaban F., Karakus F., Arslan K., Cinar M.U., Akyuz B. The Role of *DGAT1-Alul* and *CAST-MspI* Polymorphisms in Early Growth Traits of Morkaraman Lambs. *Veterinary Medicine and Science*. 2025; 11(3): e70395. <https://doi.org/10.1002/vms3.70395>

39. Zhang Y. *et al.* Association of *BMP15* and *GDF9* Gene Polymorphisms with Litter Size in Hu Sheep. *Genes*. 2025; 16(2): 168. <https://doi.org/10.3390/genes16020168>

40. Budiyo A., Hartanto S., Widayanti R., Setyawan E.M.N., Wardono H.P., Gustari S. The relationship between G1 (c. 260 G > A) and G4 (c. 721 G > A) polymorphisms in the *GDF9* gene and the litter size of sheep: A meta-analysis study. *Journal of Advanced Veterinary and Animal Research*. 2023; 10(4): 599–607. <http://doi.org/10.5455/javar.2023.j715>

41. Оздемиров А.А., Чижова Л.Н., Хожокоев А.А., Суржикова Е.С., Догеев Г.Д., Абдулмагомедов С.Ш. Полиморфизм генов *CAST*, *GH*, *GDF9* овец дагестанской горной породы. *Юг России: экология, развитие*. 2021; 16(2): 39–44. <https://doi.org/10.18470/1992-1098-2021-2-39-44>

42. Volkova V.V. *et al.* Investigation of the Genetic Diversity of Dagestan Mountain Cattle Using STR-Markers. *Diversity*. 2022; 14(7): 569. <https://doi.org/10.3390/d14070569>

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